

Male-specific (*Z*)-9-tricosene stimulates female mating behaviour in the spider *Pholcus beijingensis*

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Chemical signals play an important role in spider sexual communication, yet the chemistry of spider sex pheromones remains poorly understood. Chemical identification of male-produced pheromone-mediating sexual behaviour in spiders has also, to our knowledge, not been reported before. This study aimed to examine whether chemically mediated strategies are used by males of the spider *Pholcus beijingensis* for increasing the probability of copulation. Based on data from gas chromatography–mass spectrometry analysis, electroantennography assay and a series of behavioural tests, we verified that (*Z*)-9-tricosene is a male-specific compound in the spider *P. beijingensis*. This compound acts as an aphrodisiac: it increases the likelihood that a female will mate. Mate-searching males release (*Z*)-9-tricosene to stimulate sexual behaviour of conspecific females. In the two-choice assay, however, sexually receptive females show no preference to the chambers containing (*Z*)-9-tricosene. This indicates that the male pheromone of *P. beijingensis* is not an attractant *per se* to the conspecific females. This is, to our knowledge, the first identification of a male-produced aphrodisiac pheromone in spiders.

Keywords: (*Z*)-9-tricosene; male pheromone; aphrodisiac; Araneae; Pholcidae

1. INTRODUCTION

Accurate mate identification is an important component in spider sexual communication. The exchange of chemical signals is probably the first type of communication in spiders that serves to bring males and females together (Weygoldt 1977). Although chemical communication has long been recognized in spiders, most of the early research concentrated on behavioural assays of spider sex pheromones (Prenter *et al.* 1994; Searcy *et al.* 1999; Anderson & Morse 2001; Kasumovic & Andrade 2004; Roberts & Uetz 2005; Leonard & Morse 2006; Stoltz *et al.* 2007). The chemistry of spider pheromones remains poorly understood (Gaskett 2007). Some studies have investigated the chemistry of spider pheromones, and a few sex pheromones of females have been identified to date (Schulz & Toft 1993; Prouvost *et al.* 1999; Papke *et al.* 2000, 2001; Tralalon *et al.* 2005; Xiao *et al.* 2009; Chinta *et al.* 2010; Jerhot *et al.* 2010). Although all the identified spider pheromones are released by females and received by males, there are known bioassay examples of male pheromones in spiders that mediate the courtship behaviour of conspecific males or females. For example, silk extracts from mature males of the wolf spider, *Schizocosa ocreata*, can affect the frequency of agonistic displays and inhibit courtship behaviour among conspecific males. This is indicative of male-produced pheromones bound to the silk acting as a male–male inhibitor (Rao Ayyagari & Tietjen 1987). In the funnel-web spider, *Agelenopsis aperta*, all females enter a

quiescent state prior to the initiation of mating (Singer *et al.* 2000). The males can complete mating when the female is physiologically or behaviourally inactive. Becker *et al.* (2005) demonstrated that the courting males emit an airborne pheromone to induce female quiescence during courtship. Males of a small theridiid spider (*Argyrodes* sp.) have a protuberance on the front of their heads. This secretes an aphrodisiac that is sucked on by females during mating (Legendre & Lopez 1974; Becker *et al.* 2005). No one, to our knowledge, has reported, however, a chemical identification of male-produced pheromone-mediating sexual behaviours in spiders.

Pholcid spiders (Araneae, Pholcidae), known colloquially as ‘daddy-long-leg spiders’, are among the dominant web-building spiders distributed worldwide. They occupy a wide variety of habitats ranging from leaf litter to tree canopies; several species occur in caves and in close proximity to humans (Huber 2005). Sexual selection by female choice occurs in pholcids (Uhl 1998; Huber 1999; Uhl *et al.* 2005). *Pholcus beijingensis* is a common species found in various caves near Beijing in China. Spiders of this species usually construct untidy webs in dark and damp recesses of cave entrances (Chen & Li 2005). *Pholcus beijingensis* is polygamous and males and females have multiple mating partners. Males abandon their webs after their last moult to search for potential mates while the females wait on their webs for males (Chen & Li 2005). We have demonstrated that a combination of (*E,E*)-farnesyl acetate and hexadecyl acetate acts as a female-produced sex pheromone in this species (Xiao *et al.* 2009). The mate-searching males (MMs) can locate potential mates based on the sex pheromone

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associated with the female's silk. The approach of an MM, unlike a female or immature male, rarely triggers predatory or aggressive behaviour in the sexually receptive female (Chen & Li 2005; Xiao *et al.* 2009). Therefore, we hypothesized that MMs might emit scents acting at a close range as sexual attractants and/or stimulants for the conspecific mature females. To test the hypothesis that a male-produced pheromone exists in *P. beijingensis*, we isolated and identified chemical signals from the body extract of *P. beijingensis* and tested for chemoreceptor responses to the compound using electroantennography. We confirmed behavioural responses with female choice tests and behavioural assays.

2. MATERIAL AND METHODS

(a) Spiders

Adult and subadult specimens of *P. beijingensis* were collected in March and April of 2008 at the entrance of a cave located to the southwest of Beijing (39°42.350' N, 115°42.825' E). Mean body weights of the adult males, adult females and subadult males (SMs) were 18.2 ± 3.3 mg ($n = 10$), 14.6 ± 2.8 mg ($n = 10$) and 9.6 ± 1.7 mg ($n = 12$), respectively. Each spider was kept in a glass cuvette (4 cm inner diameter (i.d.) 12 cm deep) with a small moistened wad of cotton on the bottom to provide humidity. The cuvettes were put in a climatic chamber (RXZ-268B, Ningbo Jiangnan Instrument Factory) under a 12 L:12 D photoperiod regime at 25°C (day) and 23°C (night). About 10–15 fruitflies (*Drosophila melanogaster*) were provided to each spider for food once a week.

(b) Chemical procedures

As the site of pheromone production in the spider is unknown, we extracted the whole body for gas chromatography–mass spectrometry (GC–MS) analysis. Whole-body extraction was carried out as described (Budenberg *et al.* 1993; Leal *et al.* 1994). Sexually receptive females (RFs), MMs and SMs supplied the body extraction samples. In courtship sequences of *P. beijingensis*, the male unfolding its pedipalps towards the female is seen as a critical behavioural pattern because it implies courting success. At this point, the female turns from passive to active and approaches the male for mating (Xiao *et al.* 2009). To determine whether the adults were reproductively active, we paired each male with an adult female on her web and checked for courtship behaviour. We removed the male from the web when it unfolded its pedipalps towards the female. Ten spider pairs from the total 15 pairs were found to be sexually receptive in a 1 h observation period. These sexually RFs and males were chosen for the chemical analysis. Each spider was put into a 100 µl cuvette, which was inserted into a 1.5 ml screw-topped vial (Agilent Technologies, Santa Clara, CA, USA). Then, 40 µl dichloromethane (purity greater than 99.5%, Beijing Fine Chemical Company, Ltd, Beijing, China) was put into the cuvette to extract compounds from the spider body. Twenty-four hours later, we removed the spider from the cuvette and stored the remaining solution at –20°C until analysis by GC–MS.

Analytical GC–MS was performed on an Agilent Technologies Network 6890N GC system coupled with a 5973 Mass Selective Detector using the NIST/EPA/NIH Mass Spectral Library (2002 version; Agilent Technologies). Chemstation software (Windows 2000) was used for data

acquisition and processing. The GC was equipped with a 30 m HP5 – MS capillary column (0.25 mm i.d. × 0.25 µm film thickness; Agilent Technologies). Helium was used as the carrier gas at a flow rate of 1.0 ml min⁻¹. The temperature of the injector was set at 280°C. Two microlitre aliquots of each sample were injected in the splitless mode. As dichloromethane is very volatile, we measured the volume of the solution remaining from the body extracts after injecting 2 µl into the GC–MS instrument so that we could calibrate the proportion of the extract injected for titre analysis. The oven temperature was programmed to increase from 100 to 300°C at 5°C min⁻¹ and then held for 15 min. Electron-impact ionization used 70 eV and the scanning mass ranged from 30 to 450 amu. Compounds were identified tentatively by matching their gas chromatographic retention times and mass spectra with authentic analogues of the mass spectral library. Synthetic (*Z*)-9-tricosene (purity greater than 99.4%; Tokyo Chemical Industry Co., Ltd, Tokyo, Japan) and tricosane (purity greater than 99%, Alfa Aesar, a Johnson Matthey Co., MA, USA) were used to confirm the identification of natural products after separation on a non-polar column (HP5 – MS, 0.25 mm × 30 m × 0.25 µm) and a polar column (DB – wax, 0.25 mm × 30 m × 0.25 µm, J&W Scientific, Folsom, CA, USA).

We determined the content of (*Z*)-9-tricosene from the body extract of one male *P. beijingensis* using an external standard. The synthetic compound (*Z*)-9-tricosene was diluted sequentially in dichloromethane to concentrations of 0.001, 0.01 and 0.1 µg µl⁻¹. We injected 1 µl aliquots of each prepared solution into the GC–MS instrument. By comparing peak areas of the body extracts and those of the (*Z*)-9-tricosene solutions, we estimated that the peak areas of most male body extracts were larger than that of 0.001 µg (*Z*)-9-tricosene standard, but smaller than that of 0.01 µg. Therefore, we diluted (*Z*)-9-tricosene in dichloromethane to concentrations closely similar to the component of the male body extract for titre analysis. We injected 0.001, 0.002, 0.003, 0.004, 0.005, 0.006, 0.008 and 0.01 µg of the synthetic sample to obtain a calibration regression equation. The quantity of the male-produced (*Z*)-9-tricosene from one male body extract was calculated by comparing the peak area with that of the synthetic standard sample. We calculated the quantity of the male-produced pheromone from one male body extract according to the calibration regression equation and the volume of the extract solution.

(c) Electroantennogram (EAG) recording

The olfactory chemoreceptors for airborne, volatile pheromones have not yet been determined in spiders (Foelix 1996). Odours of conspecific females and of prey species, however, evoke electrical reactions in male pedipalps (Gemeno *et al.* 2000; Tichy *et al.* 2001). Putative chemoreceptors on female spider legs have been examined by electron microscopy (Ross & Smith 1979; Barth 2002). Electroantennogram (EAG) responses to the synthetic tricosane and (*Z*)-9-tricosene were recorded from the first leg tarsus of sexually RFs in our experiment. One leg of the tested spider was excised at the proximal base of the tarsus. The tip of the terminal tarsus segment was removed. Both the proximal end and distal ends of the tarsus were stuck on the electrode holder (PRG-2 probe; Syntech, Hilversum, The Netherlands) using electrically conductive gel. The electrode holder was connected to a high-impedance signal amplifier (Intelligent Data Acquisition Controller CS-55,

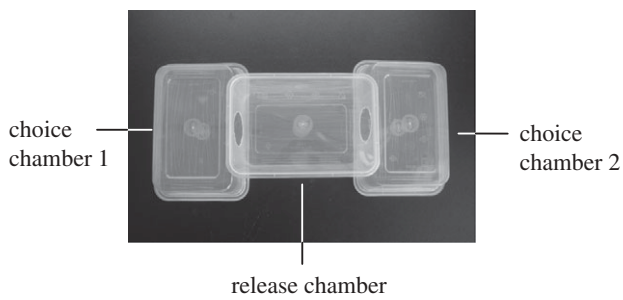


Figure 1. Two-choice arena system used in the behavioural assays. The three chambers were upended to each other and spliced with a hole on one or two sides. The selecting spider was released in the central chamber and could move freely to the left or right choice chamber.

Syntech). The amplified electrical potential signal from the antenna was recorded on a personal computer using the Syntech EAG software. An odour puff filtered through an active carbon filter was mixed with a humidified air-stream blowing continuously over the tarsus preparation at a rate of 250 ml min^{-1} through a silicone rubber pipe connected to a glass tube terminating 3 mm from the tarsus preparation. Both tricosane and (*Z*)-9-tricosene were dissolved in dichloromethane at $0.1 \mu\text{g } \mu\text{l}^{-1}$. Ten microlitre aliquots of each tested compound or dichloromethane alone (control) were absorbed on pieces of filter paper ($5 \times 30 \text{ mm}$). The filter paper was inserted into the glass pipette and exposed to the air stream 8 min later, by which time the solvent had evaporated at room temperature (25°C). The order of the odour stimuli was as follows: air (control), dichloromethane (solvent blank), tricosane, (*Z*)-9-tricosene, dichloromethane and air. The EAG responses were tested using five tarsus preparations each of a female *P. beijingensis*. All tests were repeated twice for each tarsus.

(d) Behavioural assays

First, we tested the attractiveness of (*Z*)-9-tricosene to sexually RFs in a two-choice arena system used in previous behavioural assays of male responses to the female-produced sex pheromone of *P. beijingensis* (Xiao *et al.* 2009). The three chambers were upended and spliced with a hole on one or two sides (figure 1). The test female was released in the central chamber and could move freely to the left or right choice chamber. Sexually RFs were separated into three groups to test the attractiveness of (*Z*)-9-tricosene, tricosane, and a blend of (*Z*)-9-tricosene and tricosane, respectively. (*Z*)-9-tricosene, tricosane and the binary blend were dissolved in dichloromethane at concentrations of 0.001, 0.01 and $0.1 \mu\text{g } \mu\text{l}^{-1}$. The test females were introduced into the central release chamber and allowed to acclimatize to the surroundings for 1 h before the trials. A piece of filter paper containing $10 \mu\text{l}$ of the test solution was placed in one of the choice chambers; the other choice chamber contained a filter paper with an equal quantity of dichloromethane (solvent control). The choice chambers contained the chemical solution or the control solvent alternately between successive trials to eliminate any possible bias. As searching behaviour of this spider only occurs in the dark (Y.-H. Xiao, J.-X. Zhang & S.-Q. Li 2008, unpublished data), these tests were completed at night (usually from 20.00 to 22.00 h). We observed movement of the test females under infrared light and recorded which chamber it chose first during a 2 h observation period. If any test female was

disturbed and escaped into a choice chamber as soon as the choice arena system was connected, we discarded the trial. Each test female was used twice in the two-choice behavioural assay. We rested each female for one week before the second test trial.

Second, we tested whether the sexually RFs could detect (*Z*)-9-tricosene as a cue of gender from the conspecific males and thus adjust their behaviour to mate with the males. In the laboratory experiments, males always held still on or under the female webs for a short time (usually from a few minutes to half an hour) after they had been introduced into the female webs. After a short interval for acclimation, the males started to move on the web to search for the female, which is the beginning of male courting sequences. As our former study mentioned (Xiao *et al.* 2009), male spiders of *P. beijingensis* display courting behaviour actively during most of the courtship sequence while females stay motionless on the web. When the male unfolds its pedipalps, the female turns from passive to active and approaches the male for mating. If females were not sexually receptive (e.g. subadult females or gravid adults), they would attack the intruders as soon as we introduced the male spiders on their webs. In a sense, keeping motionless (instead of showing aggressive behaviour) indicates the sexual acceptability of the females. Therefore, we compared male acclimation time, male courting time and mating time to illustrate differences between the treatment groups and the control group. Square boxes made of plastic-coated cardboard ($22 \times 22 \text{ cm}$; 8.5 cm deep) with transparent glass tops were used for observing spider courtship and copulation. All boxes were cleaned with 95 per cent ethanol and air dried before use. A glass dish with cotton soaked in distilled water was set in each box to supply humidity. Sexually RFs were randomly divided into four groups to observe the courting and mating behaviour of the males while exposed to (*Z*)-9-tricosene, tricosane, binary blends of (*Z*)-9-tricosene and tricosane, and dichloromethane (solvent control), respectively. Females were released into the box individually and were allowed to build their webs for 24 h prior to the trials. We placed a piece of filter paper with $10 \mu\text{l}$ of test solution at $0.1 \mu\text{g } \mu\text{l}^{-1}$ in each treated female box, while a filter paper with $10 \mu\text{l}$ dichloromethane alone was placed in each control female box. Twenty minutes later, MM spiders were introduced into all treated and control female boxes (one for each). For each trial, we recorded three parameters. (i) Male acclimating time, which is defined as the interval (min) between the male introduction into the female box and the male first moving on the web. During this period, the male holds still on or under the female web to acclimate to the surroundings. (ii) Male courting time, which is defined as the interval (min) between the male first moving on the web and mating initiation. (iii) Mating time, which is defined as the interval (min) between the male inserting his pedipalps into the female's genital pore until the mates disengaged. We observed 10 pairs in each treatment group and in the control group. Three pairs in the tricosane group and one pair in each other group failed to mate. Each female and male was used only once.

Third, we tested whether the sexually RFs could detect (*Z*)-9-tricosene as a cue for the presence of prey, and adjust their behaviour to take aggressive action against the intruders efficiently. All test females were kept individually in a glass cuvette (4 cm i.d. \times 12 cm deep), in which they had lived for several days to acclimatize. No food was provided to the test spiders for 6 days before each trial.

The females were divided into two groups: a treatment group ($n = 17$) and a control group ($n = 17$). We placed a filter paper with 10 μl of (*Z*)-9-tricosene at 0.1 $\mu\text{g } \mu\text{l}^{-1}$ in each treated female cuvette, while a filter paper with 10 μl of dichloromethane solvent was placed in each control female cuvette. Twenty minutes later, 10 fruitflies were introduced into each treated and control female container. We observed predatory behaviour of the females for 2 h. The number of fruitflies caught by the females and the time that females spent on catching the first prey were recorded. Nine females in the test group and two females in the control group did not display predatory behaviour during the observation period. Each female was used only once.

(e) Statistical analyses

We measured the relative abundance of each compound by converting the peak area of a particular compound into a percentage of the summed peak areas from the 13 main GC peaks. First, we compared differences in relative abundance between the compounds extracted from MM and RF individuals; second, we compared quantitative differences between compounds from MM and SM spiders. These were analysed using either independent two-tailed Student's *t*-tests when the data were normally distributed or non-parametric Mann–Whitney *U*-tests when the data were not normally distributed.

Differences in EAG responses among various odour stimulus groups were analysed by one-way analysis of variance (ANOVA) with the least significant difference (LSD) test.

In the first behavioural assay, we used the χ^2 -test for goodness-of-fit to compare observed with expected counts for female choice data obtained from the two-choice experiment so that we could determine whether (*Z*)-9-tricosene or tricosane was attractive to the test females. In the second behavioural assay, the Kruskal–Wallis test was used when comparing differences in male acclimation and courting time among the four groups: the (*Z*)-9-tricosene exposure female group, the tricosane-exposure female group, the binary blend exposure female group and the control group, which did not have normally distributed raw data. Mann–Whitney *U*-tests were used for paired comparisons for courting time. Differences in the mating time among the four groups were analysed using one-way ANOVA as the raw data were normally distributed. In the third behavioural assay, differences in the time taken for catching the first prey between the (*Z*)-9-tricosene exposure female group and the solvent exposure female group were assessed using the Mann–Whitney *U*-test. Differences in the numbers of prey eaten between the treatment group and the control group were analysed using independent two-tailed Student's *t*-tests.

All statistical analyses were conducted using SPSS for Windows (v. 15.0; SPSS Inc., Chicago, IL, USA).

3. RESULTS

(a) Chemical identification

More than 20 different compounds in the whole-body extracts of *P. beijingensis* were detected by the GC–MS. Most were straight-chain aliphatic compounds, including aldehydes, ketones, acids, alkenes and alkanes. We tentatively identified 13 compounds that eluted in less than 30 min by matching GC retention times and mass spectra with analogues in the mass spectral library (figure 2). GC detection showed that compound 9 was present in body

extracts of MM spiders while absent from body extracts of RFs and SMs (table 1). Except for compounds that were qualitatively different between the body extracts of the three spider groups, we also performed quantitative analyses on relative abundances of the relevant compounds obtained from the body extracts (table 1). Compound 10 showed a significantly greater proportion in body extracts of MM spiders than that of RFs or SMs. Compounds 9 and 10 were tentatively identified as (*Z*)-9-tricosene and tricosane, respectively, according to their retention times and mass spectra and later confirmed with the synthetic sample after separation on HP5–MS and a DB–wax column. The mass spectra of these natural products matched those of synthetic standards (figure 3).

(b) EAG responses

EAG responses of first leg tarsi of RF spiders to the putative pheromone components (*Z*)-9-tricosene and tricosane are shown in figure 4. The tarsus preparations showed strong responses to (*Z*)-9-tricosene. The mean EAG amplitude response to (*Z*)-9-tricosene (0.223 mV) was significantly higher (ANOVA with LSD test, $p < 0.01$) than responses to the air blank (0.022 mV), solvent control (0.028 mV) or tricosane (0.030 mV). Nevertheless, differences in EAG responses among the tricosane-stimulating group, the air-stimulation group and the chloromethane-stimulating group are not statistically significant (LSD test, $p > 0.05$). These results clearly demonstrate that (*Z*)-9-tricosene evoked significant EAG responses from the tarsi preparations of the sexually RFs, while tricosane seemed to be inactive.

(c) Titre analyses

We determined the quantity of (*Z*)-9-tricosene from the body extract of one male by comparing the peak GC–MS area detected from the extract of an MM spider with that detected from the synthetic standard. The calibration regression equation is $y = 0.302x - 0.259$ ($r^2 = 0.918$, $S = 0.311$, $p < 0.001$). We calculated the quantity of the male-produced pheromone from one male body extract according to the calibration regression equation and the volume of the extract solution. There were large individual variations in pheromone titre from MM spiders. The amount of (*Z*)-9-tricosene varied from 0.033 to 0.342 μg , with a mean value of $0.114 \pm 0.036 \mu\text{g}$ (\pm s.e.).

(d) Behavioural responses of females

In the two-choice arena trials, sexually RFs displayed no significant preference between the treated chamber and the solvent chamber in the trials of three chemical dosages: (*Z*)-9-tricosene: χ_1^2 (0.01 μg) = 0.474, $p = 0.491$; χ_1^2 (0.1 μg) = 0.034, $p = 0.853$; χ_1^2 (1 μg) = 0.862, $p = 0.353$; tricosane: χ_1^2 (0.01 μg) = 0.391, $p = 0.532$; χ_1^2 (0.1 μg) = 0.143, $p = 0.705$; χ_1^2 (1 μg) = 0.034, $p = 0.853$; and the binary blend: χ_1^2 (0.01 μg) = 0.615, $p = 0.433$; χ_1^2 (0.1 μg) = 0.048, $p = 0.827$; χ_1^2 (1 μg) = 0.727, $p = 0.394$; figure 5). These negative results of the two-choice assay show that neither (*Z*)-9-tricosene nor tricosane is an attractant released by the male *P. beijingensis* for conspecific females.

In the second behavioural assay, there was no significant difference in male acclimation time among the

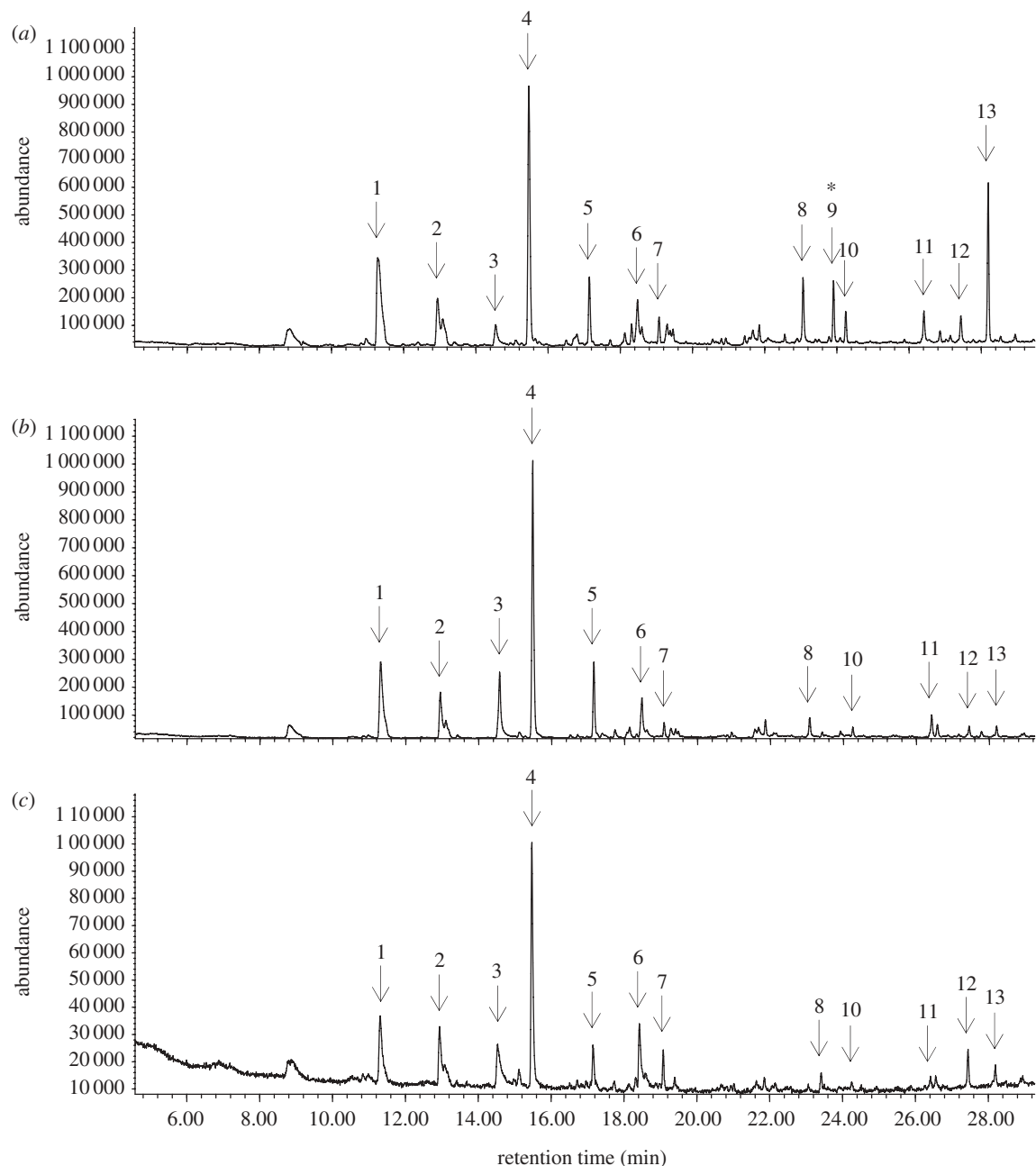


Figure 2. Total ion chromatograms of the crude extract from (a) a mate-searching male, (b) a sexually receptive female and (c) a subadult male. A non-polar column HP5-MS (0.25 mm \times 30 m \times 0.25 μ m) was used. The numbers that label the GC peaks correspond to peak numbers in table 1. The asterisk indicates peak 9 is specific in the chromatogram of mate-searching male extract.

binary blend group, the (*Z*)-9-tricosene group, the tricosane group and the control group ($\chi^2_1 = 1.473$, d.f. = 3, $p = 0.689$, Kruskal-Wallis test; figure 6). In all successful mating pairs, average mating times of the mates in the three treatment groups were a little longer than those of the mates in the control group but there was no significant difference between them ($F = 1.632$, d.f.₁ = 3, d.f.₂ = 30, $p = 0.203$ by one-way ANOVA; figure 6). Males in the binary blend group and the (*Z*)-9-tricosene group, however, spent much less time courting females than did males in the tricosane group and the control group ($\chi^2_1 = 8.407$, d.f. = 3, $p = 0.038$, Kruskal-Wallis test; figure 6). The difference of male courting time between the binary blend group and the (*Z*)-9-tricosene group was not significant ($U = 28.500$, $p = 0.288$, Mann-Whitney *U*-test). This indicates that tricosane is not a vital component for stimulating sexual behaviour in females.

In the third behavioural assay, females exposed to (*Z*)-9-tricosene spent a mean of 22.65 min in catching the first fruitfly, while females exposed to the solvent dichloromethane spent a mean of 25.93 min. There was no significant difference between these times ($p = 0.673$ by Mann-Whitney *U*-test; table 2). This result indicates that females in the treatment group did not initiate predatory behaviour more quickly than did females in the control group. On the contrary, 88 per cent of females in the control group caught the fruitflies for food while just 47 per cent of females in the treatment group caught their prey during the 2 h observation period. Females in the control group caught a mean of 2.35 fruitflies for food during the 2 h period, while females in the test group caught a mean of only 1.0. This difference was statistically significant ($t = 2.380$, d.f. = 32, $p = 0.023$, by Student's *t*-test; table 2).

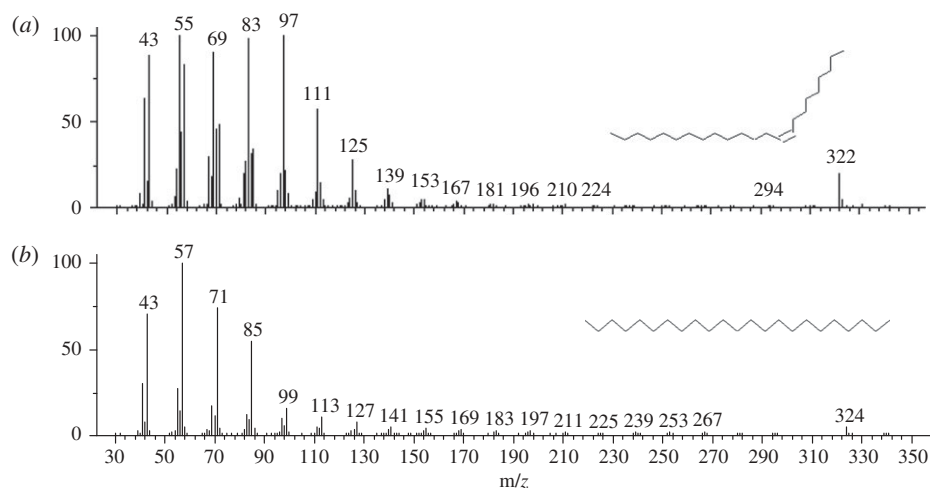


Figure 3. Mass spectra of (a) peak 9 and (b) peak 10 in the body extract of mate-searching males. They were identified as (*Z*)-9-tricosene and tricosane by comparing retention time and mass spectra with analogue in the mass spectra library (NIST 2002) and confirmed after separating the authentic standards on a non-polar column (HP5-MS) and a polar column (DB-wax).

Table 1. Comparison of relative abundance of compounds in body extracts of the spider *P. beijingensis* (mean \pm s.d.). (MM, mate-searching males; RF, sexually receptive females; SM, subadult males.)

peak no.	retention time (min)	compounds	relative abundance (%)			statistical significance (<i>p</i>)	
			MM (<i>n</i> = 8)	RF (<i>n</i> = 9)	SM (<i>n</i> = 9)	MM versus RF	MM versus SM
1	11.28	tetradecanal	19.09 \pm 3.09	22.97 \pm 3.77	10.36 \pm 5.27	0.036	0.001
2	12.94	heptadecene	4.89 \pm 1.20	6.44 \pm 1.35	5.31 \pm 2.27	0.025	0.645
3	14.54	tetradecanoic acid	4.08 \pm 4.05	6.62 \pm 5.61	10.29 \pm 4.57	0.386 ^a	0.009 ^a
4	15.46	hexadecanal	26.76 \pm 5.02	32.89 \pm 2.32	26.28 \pm 7.59	0.005	0.884
5	17.14	2-heptadecanone	6.18 \pm 1.62	6.60 \pm 1.30	3.05 \pm 1.47	0.560	0.003 ^a
6	18.48	hexadecanoic acid	4.24 \pm 3.33	4.07 \pm 2.17	12.43 \pm 4.69	0.904	0.001
7	19.08	tricosene	2.43 \pm 0.47	1.91 \pm 0.32	3.84 \pm 3.63	0.016	0.630 ^a
8 ^b	23.06	silaceous compound	3.98 \pm 1.34	3.64 \pm 1.40	4.30 \pm 2.40	0.615	0.744
9 ^{c,d}	23.91	(<i>Z</i>)-9-tricosene	4.53 \pm 2.08				
10 ^{d,e}	24.24	tricosane	4.56 \pm 2.87	1.09 \pm 0.59	2.00 \pm 0.97	0.011	0.041
11 ^b	26.41	silaceous compound	2.91 \pm 0.47	3.01 \pm 0.80	4.13 \pm 1.46	0.847 ^a	0.124 ^a
12	27.44	pentacosane	8.31 \pm 6.75	1.79 \pm 1.15	9.60 \pm 5.59	0.029	0.673
13 ^f	28.20	diisooctyl phthalate	8.04 \pm 3.34	8.98 \pm 6.14	8.40 \pm 6.70	0.697	0.891

^a*p*-values were tested by using the Mann-Whitney *U*-test; others were tested by using the independent *t*-test.

^bThe silaceous compound were not considered as compounds from the spider body extracts but contaminants from the GC column.

^cThe compound (*Z*)-9-tricosene is specific in the body extract of mate-searching males.

^dThe compound was verified with synthetic standard samples after separation with a non-polar column (HP5-MS) and a polar column (DB-wax); other compounds were identified by comparison with spectra listed in the NIST Mass Spectral Library (Agilent Technologies 2002).

^eRelative abundance of the compound tricosane in the body extract of mate-searching males was significantly more than that in body extract of sexually receptive females or subadult males.

^fThis compound is a plasticizer contaminant and probably came from the box that was used to contain the spider to build their web.

4. DISCUSSION

Our study revealed that females of *P. beijingensis* exposed to (*Z*)-9-tricosene initiated mating with the males far more quickly than did females without exposure. MM individuals thus release (*Z*)-9-tricosene acting as a sexual stimulant for conspecific females. Males of many spider species have long been known to use chemical tactics that increase the probability of mating. For example, males release silk-bound semiochemicals for attracting females or inhibiting the courtship behaviours of conspecific males (Ross & Smith 1979; Yoshida & Suzuki 1981; Roland 1984; Rao Ayyagari & Tietjen 1987; Suter *et al.* 1987; Becker *et al.* 2005). (*Z*)-9-tricosene is, however, to our knowledge, the first identified male pheromone found in spiders. Based on the results of the behavioural assay, this pheromone acts like an aphrodisiac in that it

increases the likelihood that a female will mate (figure 6). The MM individuals appear to release (*Z*)-9-tricosene to stimulate the sexual behaviour (copulation) of RF spiders so that they can mate more quickly (figure 6). It seems that, however, this aphrodisiac pheromone is not attractive to the females because the females showed no preference to the chambers containing (*Z*)-9-tricosene in the two-choice assay (figure 5). In nature, most male spiders leave their retreats or webs at maturity and start wandering around or spin their own nests right next to the potential mates (Foelix 1996). The MMs in many spider species are apparently attracted by female sex pheromones (Gaskett 2007). In contrast to males, female spiders usually apply the sit-and-wait tactic for mating. Female *P. beijingensis* remain motionless during most of the male courtship. Only after the males unfold

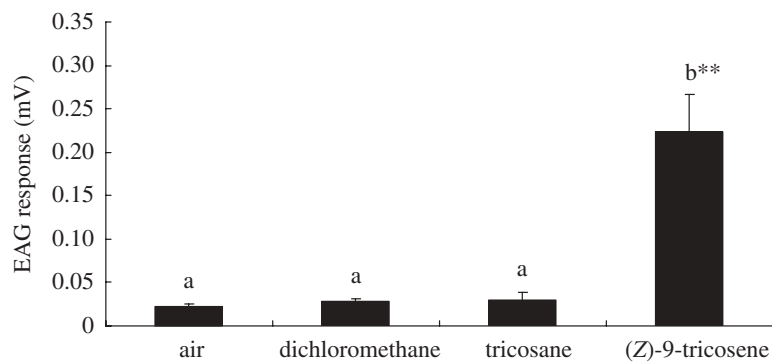


Figure 4. EAG responses (mean \pm s.e.) of first leg tarsi ($n = 5$) of sexually receptive female *P. beijingensis* to putative pheromone compounds. The order of the odour stimuli was as follows: air (control), dichloromethane (solvent blank), tricosane and (Z)-9-tricosene. ** $p < 0.01$ (one-way ANOVA with LSD test).

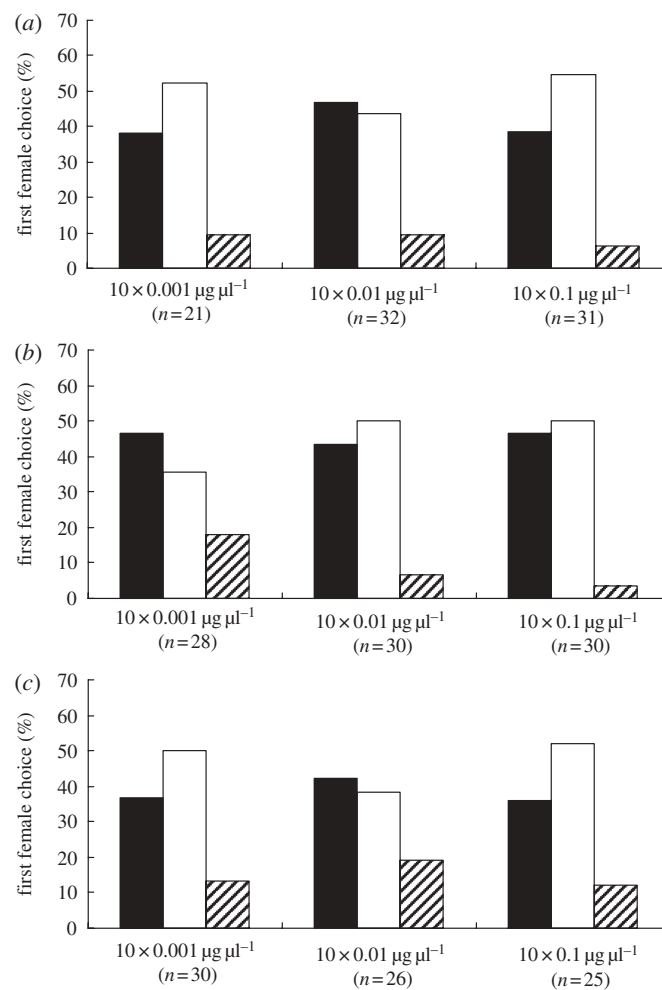


Figure 5. Results of the attractiveness for female *Pholcus beijingensis* to (a) (Z)-9-tricosene, (b) tricosane and the binary blends of (Z)-9-tricosene and (c) tricosane. Trials were completed in the two-choice arena system. (a) Black bars, (Z)-9-tricosene chamber; white bars, solvent chamber; striped bars, release chamber; (b) black bars, tricosane chamber; white bars, solvent chamber; striped bars, release chamber; (c) black bars, binary blends chamber; white bars, solvent chamber; striped bars, release chamber. 'Release chamber' refers to females that within the 2 h observation period failed to leave the central arena into which they had been introduced 1 h prior to the start of the trial.

their palps at the last phase of the courting procedure will the females turn from passive to active, which leads directly to copulation (Xiao *et al.* 2009).

(Z)-9-tricosene has also been identified as a sex pheromone released by the female housefly *Musca domestica* (Carlson *et al.* 1971), which is a prey species of *P. beijingensis*. Later, some other compounds such as (Z)-9,10-epoxytricosane, (Z)-14-tricosene-10-one and

some methyl alkanes were found to enhance the male housefly's sexual activity in combination with (Z)-9-tricosene (Uebel *et al.* 1976; Rogoff *et al.* 1980). A number of animal species use chemical camouflage to lure prey or access hosts (Dettner & Liepert 1994; Akino *et al.* 1999; Geiselhardt *et al.* 2006). A good example for chemical mimicry in the spiders is found among the bolas spiders. They usually feed exclusively

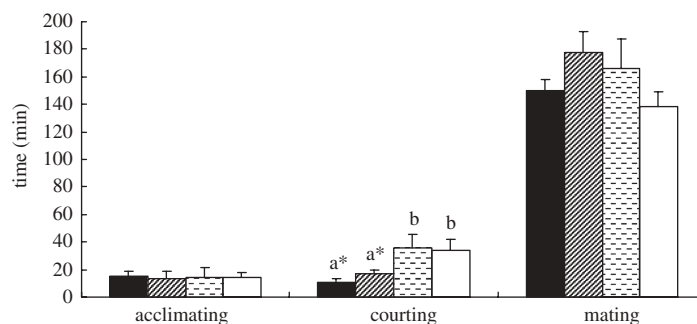


Figure 6. Influences of (*Z*)-9-tricosene, tricosane and the binary blend of both (*Z*)-9-tricosene and tricosane on courtship and mating behaviour of *P. beijingensis*. Black bars, binary blend; striped bars, (*Z*)-9-tricosene; dashed bars, tricosane; white bars, solvent control. Acclimating time is defined as the interval (min) between the male introduction into the female box and the male first moving on the web. In this period, the males keep still on the box side or bottom. Male courting time is defined as the interval (min) between the male first moving on the web and mating initiation. During this period, the male court the female until mating. Mating time is defined as the interval between the male inserting his pedipalps into the female's genital pore until the mates disengaged. Data are mean \pm s.e. * $p < 0.05$.

Table 2. Predatory behaviour of females exposed to (*Z*)-9-tricosene in comparison with that of females exposed to solvent only. Females exposed to (*Z*)-9-tricosene were in the test group and females exposed to solvent only (dichloromethane) were in the control group.

test group	predatory amount	time on catching the first fruitflies	control group	predatory amount	time on catching the first fruitflies
1	2	20	1	1	50
2	1	50	2	2	2
3	0	—	3	2	26
4	0	—	4	3	35
5	0	—	5	8	2
6	0	—	6	2	37
7	2	2	7	4	39
8	3	30	8	3	1
9	0	—	9	2	47
10	0	—	10	5	10
11	1	57	11	1	20
12	4	20	12	1	1
13	0	—	13	0	—
14	0	—	14	2	70
15	2	1	15	3	3
16	2	1	16	0	—
17	0	—	17	1	46
\bar{x}	1.00	22.65	\bar{x}	2.35 ^a	25.93

^aSignificant difference between the average number of fruitflies caught by females in the control group and those caught by females in the test group.

on males of a restricted number of moth species (Yeagan 1994). Adult female bolas spiders attract male moth prey by combinations of aggressive chemical mimicry with a specialized weapon (the bolas) and behaviour (Eberhard 1977; Stowe *et al.* 1987; Gemeno *et al.* 2000; Haynes *et al.* 2002). Cuticular hydrocarbons have also been reported as mimic chemicals in spiders. Thus, cuticular lipids of the spider *Gamasomorpha maschwitzi*, which lives in colonies of the Southeast Asian army ant *Leptogenys distinguenda*, and its host ant were virtually identical (Schulz 2004). This predatory spider acquires colony-specific cuticular hydrocarbons from their ant prey (Elgar & Allan 2004). The salticid spider *Cosmophasis bitaeniata*, which preys on the larvae of the green tree ant, *Oecophylla smaragdina*, mimics the cuticular hydrocarbon pattern of its host to avoid detection by major worker ants (Allan *et al.* 2002). In the study of Nentwig (1983), *Pholcus* showed high consumption rates of Coleoptera, Heteroptera, Hymenoptera Parasitica, Formicidae,

Lepidoptera, Neuroptera, Orthoptera and Dermaptera. In our laboratory experiments, *P. beijingensis* accepted almost all offered prey species including fruitflies, springtails, mosquitoes, houseflies, ants and even conspecific spiderlings. In field investigations, several insect species and other small arthropods including mosquitoes (Chironomidae, Tipulidae, Cecidomyiidae), moths (Gelechiidae), ants (Formicidae), bedbugs (Coreidae), houseflies (Muscidae) and pillbugs (Porcellionidae) were captured by *P. beijingensis* (Y.-H. Xiao, J.-X. Zhang & S.-Q. Li 2009, unpublished data). Whether the male *P. beijingensis* mimics the sex pheromones of their prey needs further investigation and is beyond the scope of this paper.

Hydrocarbons serve many functions in insects. They comprise a significant portion of the cuticular lipids that prevent desiccation, and are important in chemical communication (Howard & Blomquist 1982). (*Z*)-9-tricosene is a very common compound of cuticular hydrocarbons of insects in general. As well as in the housefly, (*Z*)-9-

tricosene was also found in another fly species (*Haematobia irritans*) and males contained more than females (MacLdey 1977). Intriguingly, (Z)-9-tricosene has been found in several other insects as a biologically active component. Zhang *et al.* (2003) identified five monounsaturated compounds including (Z)-9-tricosene from the Asian long-horned beetle, *Anoplophora glabripennis* and demonstrated that these compounds stimulate copulatory behaviour in males. In the social wasp, *Vespa crabro*, cuticular hydrocarbons including (Z)-9-tricosene are involved in the phenomenon of nest-mate recognition (Ruther *et al.* 2002). Thom *et al.* (2007) reported that (Z)-9-tricosene along with three other hydrocarbons could be isolated from the scent of waggle-dancing foragers of the honeybee (*Apis mellifera*). These compounds are semiochemicals, inducing worker recruitment to the food source.

It is not unique that the male spider of *P. beijingensis* shares its semiochemical with an insect species. Our previous study on the silk-bound pheromone of the spider *P. beijingensis* showed that the female sex pheromone components, (E,E)-farnesyl acetate and hexadecyl acetate, are found not only in some invertebrate species such as insects but also in mammals such as voles (Xiao *et al.* 2009). The spider pheromone 8-methyl-2-nonanone, isolated from the orb-web spider *A. aperta* (Papke *et al.* 2001), resembles a known pheromone component of the caddisfly, *Hesperophylax occidentalis* (Bjostad *et al.* 1996) and of the Asia palm weevil, *Rhynchophorus ferrugineus* (Hallett *et al.* 1993). The contact sex pheromone of *Tegenaria atrica* consists of a complex mixture of methyl esters and their fatty acids, which are semiochemicals of several insects such as the mosquito (*Aedes aegypti*) and the honeybee (Breed 1998; Ganesan *et al.* 2006). Nevertheless, two other identified spider sex pheromones, (R)-HBA and its dimer from the sheet-web spider *Linyphia triangularis* (Schulz & Toft 1993) and (S)-1,1'-dimethyl citrate from the wandering spider *Cupiennius salei* (Papke *et al.* 2000), seem to be structurally unique pheromone compounds in spiders.

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